

AD_____

Award Number: DAMD17-03-C-0040

TITLE: Smallpox Antiviral Drug

PRINCIPAL INVESTIGATOR: Dennis E. Hraby, Ph.D.

CONTRACTING ORGANIZATION: SIGA Technologies
Corvallis, OR 97333

REPORT DATE: January 2006

TYPE OF REPORT: Annual

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

REPORT DOCUMENTATION PAGE				Form Approved OMB No. 0704-0188	
Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Department of Defense, Washington Headquarters Services, Directorate for Information Operations and Reports (0704-0188), 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number. PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS.					
1. REPORT DATE (DD-MM-YYYY) 01-01-2006		2. REPORT TYPE Annual		3. DATES COVERED (From - To) 1 JAN 2005 - 31 DEC 2005	
4. TITLE AND SUBTITLE Smallpox Antiviral Drug				5a. CONTRACT NUMBER DAMD17-03-C-0040	
				5b. GRANT NUMBER	
				5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S) Dennis E. Hruby, Ph.D. E-mail: dhruby@sgph.com				5d. PROJECT NUMBER	
				5e. TASK NUMBER	
				5f. WORK UNIT NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) SIGA Technologies Corvallis, OR 97333				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012				10. SPONSOR/MONITOR'S ACRONYM(S)	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT Using a homology-based bioinformatics approach, a structural model of the vaccinia virus (VV) I7L proteinase was developed at Transtech Pharma. A unique chemical library of ~ 51,000 compounds was computationally queried to identify potential active site inhibitors. The resulting biased subset of compounds was assayed for both toxicity and the ability to inhibit the growth of VV in tissue culture cells and found to target the I7L enzyme. However, after several rounds of medicinal chemistry the chemists were not able to improve the pharmacokinetic and toxicological properties of this series of compounds. SIGA has since designed a new structural model of the I7L active site based on the Ulp1 cysteine proteinase (PDB: 1euv). Commercially available small molecule libraries that total approximately 1 million compounds are currently being queried to identify potential inhibitors. Potential active site inhibitors are being purchased and tested in the recently developed <i>in vitro</i> I7L cleavage assay. A new fluorescence anisotropy cleavage assay is under development and will be converted to a high-throughput screen if possible. SIGA has a 200,000 small molecule library that could be used for screening in a HTS cleavage assay. The final goal of this project is the identification of a potent antiviral against various pox viruses ready for preclinical development.					
15. SUBJECT TERMS Smallpox, cysteine proteinase, antiviral drug, HTS-screening, rational drug design					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT	18. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON
a. REPORT	b. ABSTRACT	c. THIS PAGE			USAMRMC
U	U	U	UU	17	19b. TELEPHONE NUMBER (include area code)

Table of Contents

Cover.....	1
SF 298.....	2
Table of Contents.....	3
Introduction.....	4
Body.....	4
Key Research Accomplishments.....	7
Reportable Outcomes.....	7
Conclusions.....	9
References.....	9
Appendices.....	10

Introduction

Smallpox virus is considered one of the most significant threats for use as a biowarfare agent. Due to complications from vaccination, mass immunization of the populace is contra-indicated. Our current research seeks to develop effective anti-poxvirus drug(s). Using vaccinia virus (VV) as a model system, the goal of our currently funded work is to determine whether the I7L cysteine proteinase or the G1L metalloproteinase encoded by VV is the poxvirus core protein proteinase (vCPP) that is essential for viral maturation and production of infectious progeny. We have recently demonstrated that the I7L cysteine proteinase is the vCPP (Byrd et al., 2002) and will continue to study the role of this enzyme during virus growth. Given this information however, we are also positioned to launch a concerted effort to identify and develop I7L inhibitors as candidate antiviral drugs. The specific goals of the experiments outlined in this report are to: 1) Over-express and purify enzymatically-active I7L proteins; 2) Develop both biochemical and tissue culture assays to measure I7L activity; 3) Utilize a combination of rational drug design and high throughput screening procedures to identify potential I7L inhibitors; and 4) To test candidate inhibitors for their ability to inhibit poxvirus replication in infected cells and appropriate animal models. Successful completion of these experiments will identify I7L inhibitors that can be advanced into pre-clinical and clinical development as antiviral drugs. Such drugs will be an essential addition to our pharmaceutical armamentarium against the deliberate or accidental introduction of a pathogenic poxvirus into our environment in order to protect members of the armed forces or the general populace.

Body

- 1) Expression and purification of vCPP (I7L).** SIGA has utilized many different systems to try to express enzymatically active I7L protein including numerous *E. coli* based expression systems, SIGA's proprietary SPEX and PLEX systems, mammalian cell expression, Baculovirus, and yeast. Some success has been seen with the Baculovirus and yeast systems so efforts are on-going with various recombinant constructs. SIGA is also utilizing Blue Sky Biotech's (www.blueskybiotech.com) protein expression expertise to see if they can express and purify active I7L enzyme from their baculovirus and/or yeast expression systems.

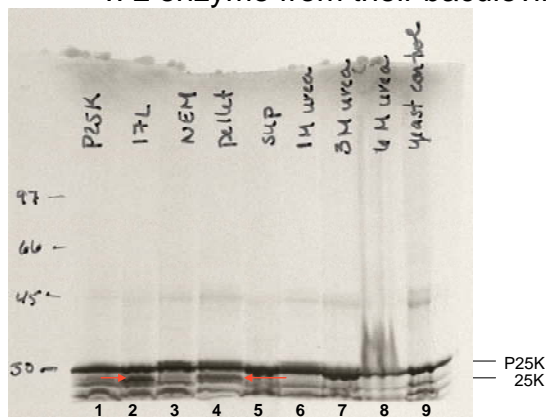
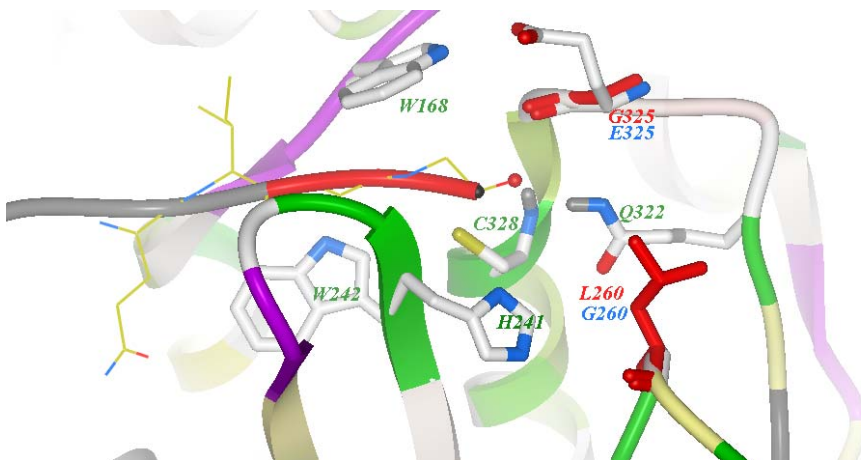


Fig 1. I7L cleavage assay. P25K made from TNT kit and 35S-met labeled. I7L from VV transient expression (lane 2-3). Lane 3 contains NEM inhibitor. I7L expressed from Yeast (lanes 4-9). Cleavage (red arrows) is seen in lane 2 (I7L from VV transient expression) and lane 4 (pellet of yeast expressing I7L).

2) **In vivo screening for inhibitor activity.** Leads identified by SIGA from TTP's library have shown specific activity against I7L and inhibit vaccinia virus replication, but the pharmacokinetic and toxicological properties are undesirable. Chemists at TTP have not been able to improve these properties through iterative chemistry. So, SIGA is using the technology at Molsoft and has built a new structural model for I7L that is shown below.

3) Rational drug design.



We have built a structural model of I7L active site based on Ulp1 cysteine proteinase (PDB: **1euv**) as the closest homologous template in PDB database. Though sequence homology is very weak (~19% identity), conservation of the key residues in the active site, as well as identical substrate specificity of Ulp1 and I7L make it possible to generate high resolution model of the binding pocket.

Indeed, our analysis shows that there is only five side chain, interacting with P2, P1 and P1' sites of the substrate, and all these five residues are identical between I7L and Ulp1 (side chains with green labels). These conserved residues include **C328** and **H241** as the active site pair, **Q322** side chain, which together with backbone amino group of **C328** forms a subtilisin-like oxyanion hole, as well as **W242** and **W168** that form a narrow channel for substrate. All the backbone contacts in this active site core are also predicted to be conserved in I7L.

Conformation of another two (non-conserved) residues, flanking the active site core from C-terminal side (P' side) can be also predicted with high accuracy (shown with blue labels for I7L and red labels for Ulp1). Thus, **E325** is predicted to form an H-bond with **W168**, while **G260** conformation is trivial.

The described active site of I7L represents a tight binding pocket with volume about 400 Å³, suitable for rational drug design. While the core of this binding pocket is fully conserved between I7L and mitochondrial protease Ulp1 in human, difference in non-conserved residues E325 and G260 should be sufficient to allow design of I7L selective inhibitors.

SIGA is using this model to search through all the commercial compound libraries available (approximately 1 million compounds total) and will test potential inhibitors in cell culture against vaccinia virus.

3) *In vitro* cleavage assay. (See attached publication for more information). SIGA is continuing to optimize the cleavage assay using I7L from vaccinia virus infected cells and substrate (P25K, p4a, p4b) produced in a transcription and translation kit. This assay will be used to test the activity of enzyme produced from Blue Sky Biotech. This assay will also be used to test potential antiviral compounds. See figure 2 below for western blot results using this assay.

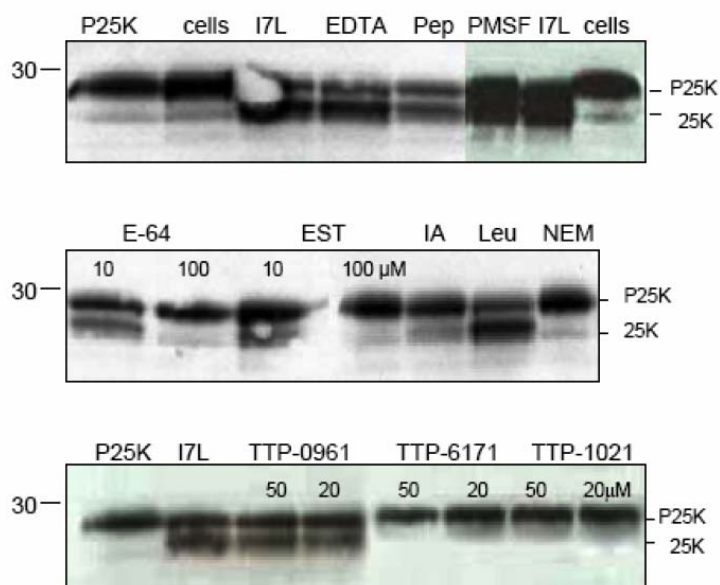


Fig 2. I7L *in vitro* cleavage assay. Substrate (P25K) mixed with various extracts (alone, cells, I7L) with or without the addition of known proteinase inhibitors and TTP compounds. If cleavage occurs you see a band at 25K.

4) Fluorescence Anisotropy assay. SIGA is continuing work on a new cleavage assay that can be done in high throughput instead of by western blot. The substrate (P25K) will be tagged with a tetraCys motif at the N-terminus and then expressed and purified from *E. coli*. I7L enzyme will be produced as in the assay above or other available system(s). For the assay the purified substrate is mixed with Lumio reagent (fluorescent reagent that binds to the tetraCys motif) and then I7L enzyme +/- inhibitors are added. Then measure fluorescence – this can be done in a 96-well plate for HTS.

5) High-throughput Screening. If the cleavage assay can be optimized for high-throughput screening, SIGA will screen through a small molecule

compound library that it just purchased from ChemBridge (~160,000 compounds) as well as a small library that SIGA already owned (36,000 compounds).

Key Research Accomplishments

- ☐ Discovery of a series of compounds that target the I7L protease and are effective against various pox viruses in cell culture
- ☐ Vaccinia virus murine intranasal challenge model set up and validated at SIGA
- ☐ Vaccinia virus murine tail vein model set up and validated at SIGA
- ☐ New rational model of I7L active site finished
- ☐ Ability to screen larger libraries (~1 million compounds) *in silico* using the new I7L model – screening in process
- ☐ Developed new I7L *in vitro* cleavage assay – published in Virology

Reportable Outcomes (manuscripts, abstracts, presentations, patents, etc)

• Manuscripts

Byrd, C.M., T.C. Bolken, and D.E. Hruby. 2002. The vaccinia virus I7L gene product is the viral core protein proteinase. J. Virol. 76:8973-8976.

Byrd, C.M., T.C. Bolken, and D.E. Hruby, 2003. Molecular dissection of the vaccinia virus I7L core protein proteinase. J. Virol 77:11279-11283.

Byrd, C.M., T.C. Bolken, and D.E. Hruby. 2003. Vaccinia virus cysteine proteinase. In: Handbook of Proteolytic Enzymes, Vol. 1-2. 2nd Edn. (Barrett,A., Rawlings,N., & Woessner, J., eds) Amsterdam: Elsevier.

Byrd, C.M., T.C. Bolken, and D.E. Hruby. 2003. Vaccinia virus proteinase. In: Handbook of Proteolytic Enzymes, Vol. 1-2. 2nd Edn. (Barrett,A., Rawlings,N., & Woessner, J., eds) Amsterdam: Elsevier.

Byrd, C. M., T. C. Bolken, A. M. Mjalli, M. N. Arimilli, R. C. Andrews, R. Rothlein, T. Andrea, M. Rao, K. L. Owens and D. E. Hruby. 2004. New Class of Orthopoxvirus Antiviral Drugs That Block Viral Maturation. J. Virol 78:12147-12156.

Byrd, C.M., and D.E. Hruby. 2005. A conditional-lethal vaccinia virus mutant demonstrates that the I7L gene product is required for virion morphogenesis. Virology Journal 2:4.

Byrd, C.M., and D.E. Hruby. 2005. Development of an *in vitro* cleavage assay system to examine Vaccinia virus I7L cysteine proteinase activity. Virology Journal 2:63.

Hruby, D.E., and C.M. Byrd. 2005. Less is more: essential role of poxvirus proteinases during assembly. ASM News Feature Article (in press).

Byrd, C.M., and D.E. Hruby. 2005. Vaccinia virus proteolysis – a review. Reviews in Medical Virology (submitted).

- **Abstracts & Presentations**

Byrd, C., S.G. Hansen, L. Hall, T. Wright, T. C. Bolken, C.H. Jones, and D.E. Hruby. 2001. Development of poxvirus proteinase inhibitors. ASV Meeting, Madison, WI. July 21-25.

Hruby, D.E., 2002. A tale of two proteinases. NIH Orthopoxvirus Working Group Annual Meeting, Washington, DC, May 14-17.

Hedengren-Olcott, M., C. M. Byrd, N. Covarrubias, and D. E. Hruby. 2002. Characterization of the VV metalloproteinase G1L. XIV International Poxvirus & Iridovirus Workshop, Lake Placid, New York, September 19-25.

Byrd, C. M., T. C. Bolken, and D. E. Hruby. 2002. Evidence that the I7L ORF encodes vaccinia virus core protein proteinase. XIV International Poxvirus & Iridovirus Workshop, Lake Placid, New York, September 19-25.

Byrd, C.M., T. Bolken and D.E. Hruby. 2003. The vaccinia virus I7L core protein proteinase as an antiviral drug target. ASM Biodefense Meeting, Baltimore, MD, March 9-12.

Hruby, D.E., C.M. Byrd, and T.C. Bolken. 2004. vvl7L: An Antiviral Drug Target. ICAR Meeting, Tucson, Arizona, May 2-6.

Honeychurch, K.M., C.M. Byrd, T. Bolken, B. Luttge, R.W. Moyer and D.E. Hruby. 2004. Characterization of novel poxvirus anti-viral targets using small molecule inhibitors of vaccinia virus replication. ASV Meeting, Montreal, Quebec, Canada, July 10-14.

Bolken, T.C., C.M. Byrd, A.M. Mjalli, M.N. Arimilli, R.C. Andrews, R. Rothlein, T. Andrea, M. Rao, M. Guzel, M. Bondlela, J. Cooper, S.S. Kickner, L.E. Sperzel, and D.E. Hruby. 2004. Optimization of Orthopoxvirus Antiviral Drugs That Block Viral Maturation. *XVth International Poxvirus and Iridovirus Symposium*. Oxford, UK, September 3-8.

Byrd, C.M. and D.E. Hruby. 2005. Smallpox antivirals: Development of and I7L in vitro cleavage assay to study core protein maturation pathways. American Society for Virology, Penn State University, PA, June 19-22.

Byrd, C.M., and D.E. Hruby. 2005. Smallpox antivirals: *In vitro* assay for Vaccinia virus I7L enzymatic cleavage of core protein precursors. International Conference on Antiviral Research, Barcelona, Spain, April 10-14.

- **Patents**

Screening Method for Orthopoxvirus Antivirals, (US & PCT applications)-preparing a Continuation in Part (CIP) to add the new I7L cleavage assays recently developed by SIGA.

Conclusions

By using homology-based computational modeling, a structural model of the VV I7L cysteine proteinase, which is responsible for essential morphogenic cleavage reactions during viral maturation, has been developed. This model is currently being used in concert with *in silico* drug docking procedures to query all commercially available chemical libraries which consist of approximately 1 million compounds total. Hits generated from this screen that are predicted to bind to the I7L catalytic site are being purchased and tested in the *in vitro* I7L cleavage assay. So far several potential inhibitors have been identified through this approach.

A cell-free transcription and translation system was used to develop an *in vitro* cleavage assay for the VV cysteine proteinase I7L. Proteolytic activity was obtained by co-expression of I7L in *ts16* infected cells at the non-permissive temperature. Each of the major core protein precursors, P4a, P4b, and P25K, were shown to be cleaved to their mature products by I7L using the *in vitro* assay. Evidence that this cleavage is specific to I7L was shown through the fact that expressing a mutant form of I7L resulted in the inability to cleave the core protein precursors. This assay will be useful in determining the activity of potential inhibitors.

SIGA is also developing a Fluorescence Anisotropy assay which could potentially be used in a high-throughput screening endeavor. SIGA recently purchased a small molecule compound library and currently has approximately 200,000 compounds that could be tested in a HTS assay.

The final goal of this project is the identification of a potent antiviral against various pox viruses ready for preclinical development.

References

1. **Andres G., A. Alejo, C. Simon-Mateo, and M. L. Salas.** 2001. African Swine Fever Virus Protease, a new viral member of the SUMO-1 specific protease family. *J. Biol. Chem* **276**: 780-787
2. **Byrd, C. M., T. C. Bolken, and D. E. Hruby.** 2002. The vaccinia virus I7L gene product is the core protein proteinase. *J Virol.* **76**:8973-6
3. **Kern, E. R.** 2003. In vitro activity of potential anti-poxvirus agents. *Antiviral Res.* **57**:35-40.
4. **Li, S. J., and M. Hochstrasser.** 1999. A new protease required for cell-cycle progression in yeast. *Nature* **398**: 246-51
5. **De Clercq, E.** 2002. Cidofovir in the treatment of poxvirus infections. *Antiviral Res* **55**: 1-13

Research

Open Access

Development of an *in vitro* cleavage assay system to examine vaccinia virus I7L cysteine proteinase activity

Chelsea M Byrd¹ and Dennis E Hruby*^{1,2}

Address: ¹Molecular and Cellular Biology Program, Oregon State University, 220 Nash Hall, Corvallis, Oregon, 97331, USA and ²Siga Technologies, 4575 SW Research Way, Suite 230, Corvallis, Oregon, 97333, USA

Email: Chelsea M Byrd - cbyrd@sgph.com; Dennis E Hruby* - dhruby@sgph.com

* Corresponding author

Published: 16 August 2005

Received: 21 April 2005

Virology Journal 2005, **2**:63 doi:10.1186/1743-422X-2-63

Accepted: 16 August 2005

This article is available from: <http://www.virologyj.com/content/2/1/63>

© 2005 Byrd and Hruby; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Through the use of transient expression assays and directed genetics, the vaccinia virus (VV) I7L gene product has been implicated as the major maturational proteinase required for viral core protein cleavage to occur during virion assembly. To confirm this hypothesis and to enable a biochemical examination of the I7L cysteine proteinase, an *in vitro* cleavage assay was developed. Using extracts of VV infected cells as the source of enzyme, reaction conditions were developed which allowed accurate and efficient cleavage of exogenously added core protein precursors (P4a, P4b and P25K). The cleavage reaction proceeded in a time-dependent manner and was optimal when incubated at 25°C. I7L-mediated cleavage was not affected by selected inhibitors of metalloproteinases, aspartic acid proteinases or serine proteinases (EDTA, pepstatin, and PMSF, respectively), but was sensitive to several general cysteine proteinase inhibitors (E-64, EST, Iodoacetic acid, and NEM) as well as the I7L active site inhibitor TTP-6171 [C. Byrd *et al.*, *J. Virol.* 78:12147–12156 (2004)]. Finally, in antibody pull down experiments, it could be demonstrated that monospecific α I7L serum depleted the enzyme activity whereas control sera including α GIL, directed against the VV metalloproteinase, did not. Taken together, these data provide biochemical evidence that I7L is a cysteine proteinase which is directly involved in VV core protein cleavage. Furthermore, establishment of this I7L-mediated *in vitro* cleavage assay should enable future studies into the enzymology and co-factor requirements of the proteolysis reaction, and facilitate antiviral drug development against this essential target.

Background

The *Orthopoxviridae* include vaccinia virus, camelpox, cowpox, ectromelia, monkeypox, raccoonpox, skunkpox, taterapox, volepox, and variola. Viruses in this family are the cause of numerous diseases including smallpox (variola), and recent human outbreaks of monkeypox. Orthopoxviruses are large double-stranded DNA viruses that are unique amongst DNA viruses in that they replicate exclusively within the cytoplasm of infected cells. Vaccinia virus (VV) is the most extensively studied virus in

this group and is the prototypic member. The genome of VV is predicted to encode over 200 open reading frames. VV expresses its genetic information in three stages, as early, intermediate, and late genes. The early genes, which account for approximately half of the genome and are transcribed prior to DNA replication, encode many of the proteins involved in viral DNA replication and intermediate gene expression. The intermediate genes, of which only a handful have been identified, are expressed after the onset of DNA replication, and encode proteins that

are activators of late gene expression. The late genes encode many proteins required for the transcription of early genes, the viral structural proteins and the enzymes necessary to process these proteins into their mature form.

Many viruses use proteolytic processing as a key step in their developmental cycle. RNA viruses and retroviruses commonly undergo formative proteolysis in which large polyproteins are cleaved by viral encoded proteinases to produce the structural and nonstructural proteins required for morphogenesis. DNA viruses such as poxviruses and adenoviruses commonly use another type of proteolysis, called morphogenic proteolysis where precursor proteins are first synthesized and then cleaved by viral proteinases to produce the mature form of the protein. The mature protein then plays an essential role in virion formation. During VV assembly, as the spherical immature virions (IVs) are maturing into the first infectious form of vaccinia virus, intracellular mature virus (IMV), a series of events takes place including proteolytic processing of viral core proteins [1-4].

Our laboratory has worked to identify and characterize the proteinases of VV in order to understand their regulation, function, and biochemistry, with a long term goal of developing inhibitors of these enzymes as antiviral drugs. The gene product of the I7L open reading frame recently has been suggested to be the core protein proteinase of VV through the use of an *in vivo trans* processing assay [5,6]. I7L is an essential late gene, as shown through temperature sensitive mutant viruses [7,8] and conditional lethal mutant viruses [9,10] where under non-permissive conditions, viral morphogenesis is blocked prior to the formation of IMV. I7L is predicted to be a 47 kDa cysteine proteinase that cleaves the major core protein precursors P4a, P4b, and P25K, products of the A10L, A3L, and L4R open reading frames respectively, at a novel Ala-Gly-Xaa cleavage site with cleavage occurring after the glycine residue [5,6]. I7L also is likely to be responsible for cleavage of the A17 membrane protein, at an Ala-Gly-Ala site [9]. This consensus Ala-Gly-Xaa cleavage site of vaccinia is similar to that used for both the adenovirus and African swine fever virus proteinases which cleave after the second glycine in a Gly-Gly-Xaa motif [11,12].

Comparative sequence analysis has suggested that the VV I7L proteinase is related to the ASFV and adenovirus cysteine proteinases and may form a new family of SUMO-1 related enzymes [13,12]. The nucleophilic cysteine is responsible for cleavage and is activated by the imidazol group of the catalytic histidine residue. Substrate specificity is determined by the substrate binding pocket and is unique for each proteinase. Several critical residues have been identified as being necessary for enzymatic activity of I7L including the catalytic triad residues [6].

Based on the identification of the catalytic residues and the predicted structure of the I7L proteinase, a new class of small molecule inhibitors was developed that are capable of inhibiting the replication of VV, and were found to specifically target I7L through the generation of drug resistant mutant viruses with the mutations mapping to I7L [14].

To date, direct studies on the enzymology of I7L-mediated proteolysis have not been possible due to the absence of a suitable biochemical assay. In the experiments reported here, we describe the development of an *in vitro* I7L-mediated cleavage assay. We have used this system to obtain both biochemical and immunological data to prove that I7L is directly involved in cleavage of the major VV core protein precursors. Having this assay available will now facilitate biochemistry of the I7L enzyme and identification of all the required reaction components to be undertaken.

Results

To date, all studies of VV I7L activity have been carried out indirectly in transfected/infected tissue culture cells. Although this approach has provided some important insights into I7L biology, it is limited with respect to the study of I7L enzymology and identification of all the *cis* and *trans* factors required for substrate identification and catalysis. In order to approach these questions, we have sought to develop an *in vitro* cleavage assay for I7L. Thus far, the obvious approaches of expressing and purifying I7L from prokaryotic and eukaryotic expression vectors and combining with peptides or proteins containing a canonical A-G-X cleavage site have not been successful (data not shown), perhaps due to either the lack of essential co-factors or inappropriate assay conditions. As an alternative approach, we sought to develop a cleavage assay using infected cell extracts as the source of I7L activity and labeled core protein precursors made *in vitro* as the substrate. If successful, this system would provide the starting point for a dissection of the essential reaction components.

In vitro Processing of Core Protein Precursors

The three major core protein precursors P4a (A10L), P4b (A3L), and P25K (L4R) which are known to be cleaved to a mature form (Figure 1) were cloned into plasmid vectors driven off of a T7 promoter to be used as a source of substrate for the assay. To investigate the ability of I7L to cleave the P4a, P4b, and P25K substrates *in vitro*, we have used a system where the substrates are produced from an *in vitro* transcription and translation assay using rabbit reticulocyte lysates and then mixed with I7L expressed from virus infected cells. BSC40 cells are infected with *ts16*, a temperature sensitive mutant virus in which the responsible mutation maps to I7L. The virus infected cells

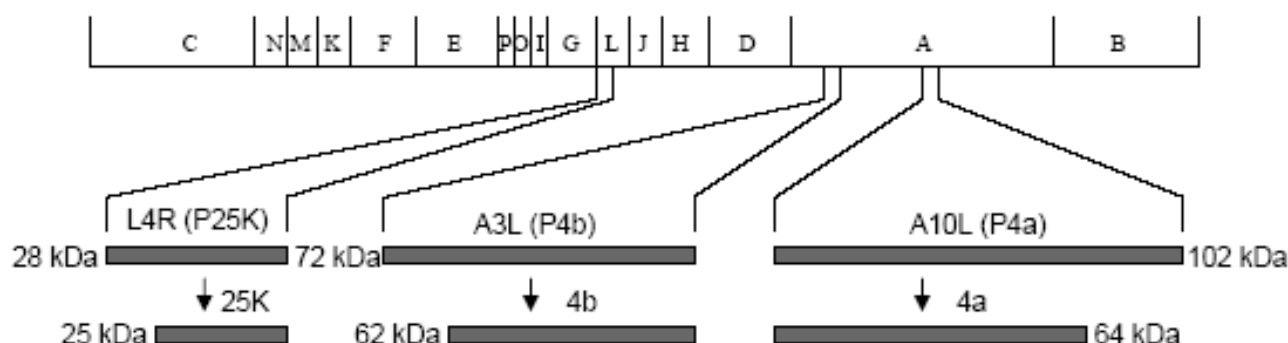


Figure 1

Schematic representation of the major core protein precursor cleavage products. The vaccinia virus genome is represented depicting three of the major core protein precursors, the gene products of the L4R, A10L, and A3L open reading frames, P25K, P4a, and P4b respectively. The precursors are shown being cleaved into their mature form. Molecular mass is indicated.

are incubated at the non-permissive temperature and transfected with plasmids expressing either wild-type I7L (pI7L) or I7L with the catalytic histidine residue mutated to an alanine (pI7LH241A). The extracts are prepared as described in the Materials and Methods. The extracts are mixed and incubated with the substrates for 3 hrs and then analyzed through SDS-PAGE and chemiluminescent detection. As shown in Figure 2, a specific band corresponding to unprocessed P4a (top panel), P4b (middle panel), or P25K (bottom panel) is produced when the substrate is run alone. When mixed with cellular extracts, or extracts from cells infected with *ts16* at the non-permissive temperature and transfected with mutant I7L, no cleavage products are observed. However, when mixed with extracts from either cells infected with *ts16* at the permissive temperature or cells infected with *ts16* at the non-permissive temperature transfected with wild-type I7L, the cleaved products 4a, 4b, and 25K are observed. Substrates with mutated A-G-X sites were not cleaved indicating that cleavage was occurring at the correct sites (data not shown). For the rest of the reported studies, P25K was used as the source of substrate since it gave the best cleavage profile.

Processing Kinetics of Core Protein Precursors

To determine the optimal temperature and kinetics of processing of the core protein precursors in the *in vitro* cleavage assay, a time course of I7L-mediated processing at various temperatures was performed. As shown in Figure 3A, at 0°C, no processing was observed during the 20 hr time period. At 25°C, a gradual increase in the amount

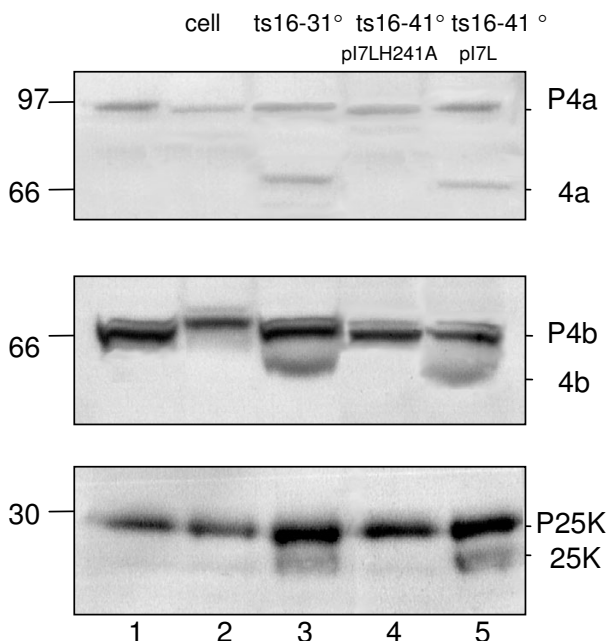
of P25K cleavage product was observed starting at 15 min and increasing throughout the 20 hr incubation period (Fig. 3B). Compared with the rate of cleavage at 25°C, cleavage was slower at 30°C (Fig. 3C), starting around 30 min and increasing through the 20 hr period, but never to the same level as at 25°C. Processing is greatly reduced at 37°C with only a faint processed band ever appearing (Fig 3D).

Influence of Thiol Reagents on the Protease Activity

Based on its sequence similarity to the adenovirus protease, the African swine fever virus protease, and an ubiquitin-degrading enzyme in yeast, as well as the identity of a catalytic triad composed of histidine, cysteine, and aspartic acid, I7L has been classified as a cysteine proteinase. The thiol reagents dithiothreitol (DTT) and cysteine have been shown to enhance the cleavage activity of the adenovirus protease in an *in vitro* peptide cleavage assay [15]. To determine whether these agents have a similar effect on the activity of I7L, they were added to the *in vitro* assay in a final concentration from 0–10 mM. However, no increase in cleavage activity was observed with the addition of either DTT or cysteine (data not shown). It is possible that once purified recombinant enzyme is produced these thiol reagents may increase its activity.

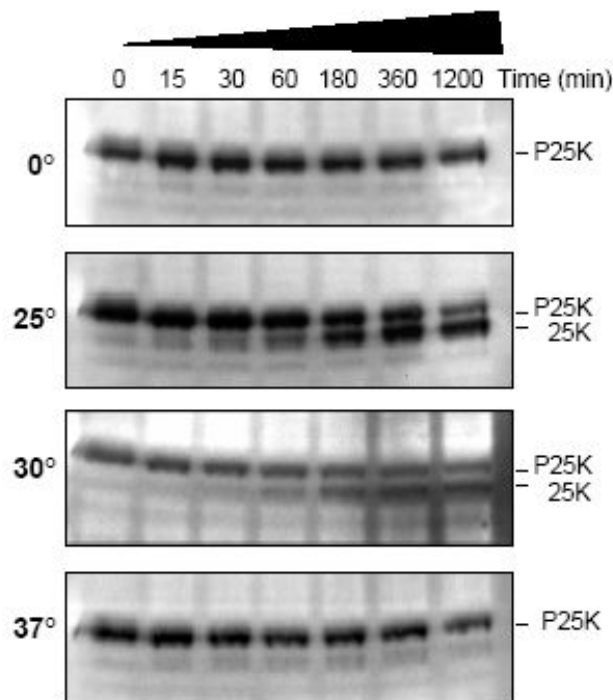
Effect of Inhibitors on Protease Activity and Characterization as a Cysteine Proteinase

The *in vitro* assay allowed us to test the effects of various protease inhibitors, as well as specific small molecule inhibitors on the activity of I7L. As shown in Figure 4 and

**Figure 2*****In vitro* proteolytic processing of P4a, P4b, and P25K.**

1 μ l of TNT produced substrate either P4a (A), P4b (B), or P25K (C) was mixed with 5 μ l of Hepes buffer and 14 μ l of enzyme extracts, either from uninfected cells, or cells infected with ts16 at the permissive or non-permissive temperature. At the non-permissive temperature, plasmid borne I7L, either wild-type (pl7L) or mutant I7L (pl7LH241A) was transfected in as the source of enzyme. The reaction was incubated at 29°C for 3 hrs before being stopped by the addition of SDS sample buffer. Molecular weight is indicated on the left and the core protein precursor and product on the right. Lane 1 is substrate alone, lane 2 is substrate mixed with cellular extracts and lanes 3–5 are substrate mixed with the enzyme extract indicated.

Table 1, the metalloproteinase inhibitor ethylenediaminetetraacetic acid (EDTA), the aspartic proteinase inhibitor pepstatin, and the serine proteinase inhibitor phenylmethanesulfonyl (PMSF) had no detectable effect on cleavage activity. The cysteine proteinase inhibitors iodoacetic acid (IA) and *N*-ethylmaleimide (NEM) efficiently blocked I7L mediated proteolysis of P25K. The cysteine proteinase inhibitors E-64 and EST were shown to inhibit protease activity at a relatively high concentration, but not at the lower concentration tested. This is consistent with what has been observed for both the adenovirus protease [16], and the African swine fever virus protease [17]. The failure of E-64 to inhibit protease activity at the lower concentration tested, and the location of the active site residues may suggest that each of these enzymes are not conventional papain-like enzymes, but

**Figure 3**

Processing kinetics of P25K. Samples were incubated at either 0°C (A), 25°C (B), 30°C (C), or 37°C (D) for up to 20 hrs, harvested at the indicated times and the reaction stopped by the addition of SDS sample buffer. Incubation temperature is indicated on the left and P25K precursor and 25K mature product are indicated on the right.

may be a new family of cysteine proteinases. The cysteine protease inhibitor leupeptin also failed to inhibit protease activity, although this lack of inhibition was also observed with the adenovirus proteinase [16].

Next we wanted to determine if the small molecule I7L inhibitors previously developed as antiviral drug candidates [14] could be shown to specifically inhibit the activity of I7L in the *in vitro* assay. The compound TTP-6171 has been shown to inhibit viral replication in tissue culture, with drug resistant virus mutations mapping to I7L [14]. Here we see that this compound along with TTP-1021, which was also found to inhibit I7L in tissue culture, inhibits the processing of P25K *in vitro*. However the compound TTP-0961, which was not found to generate resistant mutants in the I7L gene (data not shown), does not inhibit cleavage. These results demonstrate that this assay can be used for the screening of specific I7L

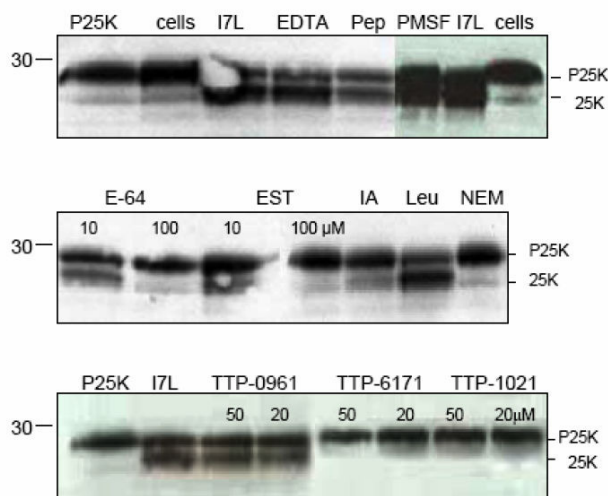


Figure 4
Effect of inhibitors on *in vitro* processing. Various concentrations of protease inhibitors were added to the *in vitro* processing assay for 6 hr at 29°C. The first lane is P25K expressed alone with no extract added. The second lane is P25K mixed with cellular extracts and the third lane is P25K mixed with I7L enzyme extracts. Each of the remaining lanes has P25K mixed with I7L enzyme extracts plus indicated inhibitor. Ethylenediaminetetraacetic acid (EDTA) was used at 1 mM. Pepstatin A, Pep, was used at 10 μM. Phenylmethanesulfonyl fluoride (PMSF) was used at 1 mM. N-(trans-Epoxy-succinyl)-L-leucine 4-guanidinobutylamide *trans*-Epoxy-succinyl-L-leucylamido(4-guanidino)butane (E-64) and a related product EST, were both used at 10 μM and 100 μM concentrations. Iodoacetic acid (IA) was used at 1 mM. Leupeptin (Leu) was used at 1 mM, and N-ethylmaleimide (NEM) was used at 2.5 mM. The concentrations of TTP-6171, TTP-1021, and TTP-0961 are indicated. The table indicates the concentration of inhibitor used and whether cleavage activity was observed.

inhibitors and confirms that this class of molecules targets I7L.

Effects of I7L antibody competition on cleavage

To directly demonstrate that the cleavage observed in the *in vitro* assay requires the presence of I7L, increasing concentrations of I7L specific antiserum were added to the enzyme extracts overnight, and then the complex was precipitated with Protein A sepharose beads to deplete the extract of I7L and any associated co-factors. As shown in Figure 5, both of the I7L antisera tested inhibited cleavage of P25K while an antiserum targeting a different VV gene product, G1L, did not inhibit cleavage.

Discussion

In this report, a cell-free transcription and translation system was used to develop an *in vitro* cleavage assay for the VV cysteine proteinase I7L. Proteolytic activity was obtained by co-expression of I7L in *ts16* infected cells at the non-permissive temperature. Each of the major core protein precursors, P4a, P4b, and P25K, were shown to be cleaved to their mature products by I7L using the *in vitro* assay. Evidence that this cleavage is specific to I7L was shown through the fact that expressing a mutant form of I7L resulted in the inability to cleave the core protein precursors. Antibody pull down experiments with αI7L supported the conclusion that I7L plays a direct role in the proteolytic reaction.

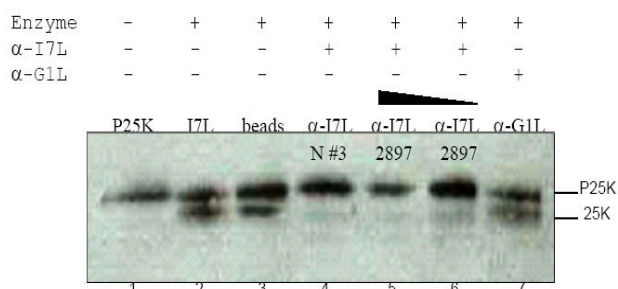
A time course of processing at various temperatures indicated that for this particular assay, the optimal temperature for the reaction to be carried out at is 25°C with processing beginning as soon as 15 minutes after addition of enzyme and increasing as time progresses. The cleavage reaction was never driven to completion and this may be due to a lack of replenishing co-factors or the enzyme may have been used up in the reaction. It was surprising that the optimal reaction temperature was 25°C instead of 37°C which is the optimal growth temperature for VV in cell culture. One possible explanation is that I7L is present at high concentrations in the extract and one can measure marginal activity at low temperature, whereas at higher temperatures other proteinases are activated which degrade the I7L enzyme.

Known cysteine protease inhibitors such as E-64, iodoacetic acid, and NEM were shown to inhibit the *in-vitro* cleavage reaction while the metalloproteinase inhibitor EDTA, the aspartic acid protease inhibitor pepstatin, and the serine protease inhibitor PMSF all failed to inhibit the cleavage reaction indicating that the enzyme responsible for cleavage is a cysteine protease. Interestingly the cysteine protease inhibitors leupeptin, and low concentrations of E-64 did not inhibit the reaction. These cysteine protease inhibitors were also not shown to be effective against either the African Swine Fever Virus protease [17] or the adenovirus protease [16], further providing support for the theory that these enzymes may form a new family of cysteine proteases that differ from papain-like cysteine proteases.

Of particular interest, the small molecule inhibitors designed to fit into the active site pocket of I7L and previously shown to inhibit viral replication [14], were found to be active in inhibiting the *in vitro* cleavage reaction described here. A related compound (TTP-0961) that was not found to map to I7L was not able to abolish cleavage. This indicated that this assay may be useful for high-

Table 1: Effect of inhibitors on *in vitro* processing.

Inhibitor	Name	Concentration	Inhibit Cleavage
Metalloproteinase	EDTA	1 mM	No
Aspartic acid proteinase	Pepstatin	10 μ M	No
Serine proteinase	PMSF	1 mM	No
Cysteine proteinase	E-64	10 μ M	No
	E-64	100 μ M	Yes
	EST	10 μ M	No
	EST	100 μ M	Yes
	IA	1 mM	Yes
	Leupeptin	1 mM	No
	NEM	2.5 mM	Yes
TTP inhibitors	TTP-6171	50 μ M	Yes
	TTP-6171	20 μ M	Yes
	TTP-1021	50 μ M	Yes
	TTP-1021	20 μ M	Yes
	TTP-0961	50 μ M	No
	TTP-0961	20 μ M	No

**Figure 5****Effect of antibody competition on *in vitro* processing.**

Lane 1 is P25K expressed alone. Lane 2 is P25K mixed with I7L enzyme extracts. Lane 3 is P25K mixed with I7L extracts that have been diluted with Hepes buffer and treated with Sepharose beads. Lanes 4, 5, and 6 are P25K mixed with I7L extracts that have been incubated overnight with different I7L antiserum (indicated on each lane), treated with Sepharose beads and the antibody complex removed by centrifugation. Lane 7 is P25K mixed with I7L extracts incubated with G1L antiserum as above.

throughput screening of compounds to identify those that have specific activity for I7L.

Conclusion

Until this point, all work demonstrating that I7L is the core protein proteinase has been done through transient-expression assays and the use of conditional lethal viruses in tissue culture [9,5,6,10]. The data obtained has indicated that I7L is essential for these processing activities, it

did not rule out the possibility that some other factor or enzyme was also required for this activity to occur. Through the use of an *in vitro* assay we have shown that I7L is capable of cleaving the core protein precursors but that an additional co-factor is required for this activity to occur since expression of the enzyme through cell-free translation produced inactive enzyme. The co-factor(s) necessary for cleavage have yet to be determined. However, having the assay described in this report available will now enable a reductive analysis to be conducted to identify all the essential components of the reaction and to study their individual biochemical characteristics.

Methods**Cells and Viruses**

BSC₄₀ cells [18] were grown in Eagle's minimal essential medium containing 5% fetal calf serum (FCS) (Sigma, St. Louis, MO), 2 mM glutamine (Invitrogen, Carlsbad, CA), and 15 μ g/ml gentamicin sulfate (Invitrogen) in a 37°C incubator with 5% CO₂. Purified ts16 Vaccinia virus was prepared as described [19]. *Escherichia coli* strains were grown in Luria-Bertani broth or on Luria-Bertani medium containing 1.5% agar and ampicillin at 50 μ g/ml.

Plasmids

The A10L (P4a) gene was amplified by polymerase chain reaction using oligonucleotides KH10 (5'-CATGCCATGGATGATGCCTATTAAGTCAATAGTTACT CTT-3') and KH11 (5'-CCGCTCGAGTTATTCATCATCAAAAGAGACAGAGTC-3'), digested with NcoI and XhoI, and cloned into the pTM1 vector, yielding pTM-P4a which utilizes a T7 promoter for expression. The A3L (P4b) gene was amplified using oligonucleotides KH08 (5'-CATGCCATGGATGGAAGCCGTGGTCAATAG-3') and KH09 (5'-

TCCCCCGGGCTAAAAATAGTTCTGTAATAT-GTCTAGCGCT-3'), digested with NcoI and SmaI, and cloned into the pTM1 vector to yield pTM-P4b. The L4R (P25K) gene was amplified using oligonucleotides DN51 (5'-CATGCCATG GATGAGTCTACTGCTAGAAAAC-3') and KH07 (5'-CCGCTCGAGTCAATCCTTT GTCG-3'), digested with NcoI and XhoI, and cloned into the pTM1 vector to yield pTM-P25K. The pI7L and pI7LH241A plasmids were described in Byrd *et al.*, 2002 [5].

Preparation of polyprotein or proteinase-containing extracts

Confluent monolayers of BSC₄₀ cells in 6-well plates were infected with ts16 VV at a multiplicity of infection of 2 plaque-forming units per cell and transfected with 2 µg of plasmid DNA (either pI7L, or pI7LH241A) using DMRIE-C (Invitrogen) following the manufacturer's indications. Infected cells were incubated either at the permissive temperature of 31.5°C or the non-permissive temperature of 39°C. Cells were harvested at 24 h post-infection by pipetting up and down to lift the cells from the surface. The infected cells were centrifuged at 10,000 × g for 10 min, the supernatant was aspirated off, and the pellet was resuspended in 500 µL homogenization buffer containing 20 mM HEPES (pH 7.4), 0.28 M sucrose, 2 mM EDTA. This was passed through a 25-gauge syringe 15 times. The homogenate was centrifuged at 700 × g for 5 min to separate the nuclei and unbroken cells from the supernatant. The supernatant was centrifuged at 100,000 × g for 30 min at 4°C to separate the membrane/particulate material from the supernatant. The supernatant was used as the source of enzyme.

Coupled TNT reactions with T7 RNA polymerase were performed according to the manufacturer's instructions (Promega Corporation, Madison, Wisconsin) as a source of substrate. Briefly, the TNT reactions were performed at 30°C in a final volume of 25 µL with 1 µg of plasmid DNA, using the non-radioactive Transcend label (biotinylated lysine residues are incorporated in the protein) provided with the kit for detection of protein.

In vitro cleavage assay

Reactions were performed at the indicated temperature in a final volume of 20 µL containing 1 µL of substrate, 13 µL of enzyme extract, and 6 µL of 20 mM HEPES (pH 7.4) buffer, pH 7.4. After the indicated times, the reaction was stopped by the addition of SDS sample buffer, and the samples were subjected to SDS-polyacrylamide gel electrophoresis. The results were analyzed by immunoblotting following the instructions provided by the TNT kit.

Inhibitor studies

For inhibitor studies, the reactions described above were incubated for 6 hr in the presence or absence of the fol-

lowing protease inhibitors: 1 mM phenylmethanesulfonyl fluoride (PMSF) (Sigma), 10 µM Pepstatin A (Sigma), 1 mM ethylenediaminetetraacetic acid (EDTA) (Sigma), 10 µM or 100 µM N-(trans-Epoxy succinyl)-L-leucine 4-guanidinobutylamide *trans*-Epoxy succinyl-L-leucylamido(4-guanidino)butane (E-64) (Sigma), 1 mM iodoacetic acid (Sigma), 10 µM or 100 µM Leupeptin (Roche, Indianapolis, IN), 2.5 mM N-ethylmaleimide (NEM) (Sigma). For I7L specific inhibition studies, the reactions described above were incubated for 6 hr in the presence or absence of TTP-6171, TTP-1021, or TTP-0961 [14] at 5 µM or 20 µM final concentrations.

Antibody competition studies

For the antibody competition studies, 25 µL of I7L or G1L specific antiserum was added to 25 µL of enzyme extract on a rotating shaker overnight at 4°C. ProteinA: Sepharose beads (Amersham Biosciences, Uppsala, Sweden) were added for 3 hrs and the antibody complex was centrifuged to pull down the I7L enzyme. The supernatant was used as the source of extract in the *in vitro* assay described above. As a control, enzyme extract was mixed with buffer instead of antibody and treated with beads in a similar manner.

Competing interests

The author(s) declare that they have no competing interests.

Authors' contributions

CMB conceived the study, conducted all the experiments and wrote the manuscript. DEH coordinated the research efforts and edited the paper. Both authors read and approved the final manuscript.

Acknowledgements

We would like to thank Kady Honeychurch for constructing pTM:L4R, pTM:A3L, and pTM:A10L, Rich Condit for providing ts16, and TransTech Pharma for supplying TTP-6171, TTP-1021, and TTP-0961. This work was funded by NIH grant AI-060160.

References

1. Moss B, Rosenblum EN: **Protein cleavage and poxvirus morphogenesis: tryptic peptide analysis of core precursors accumulated by blocking assembly with rifampicin.** *J Mol Biol* 1973, **81**:267-269.
2. Silver M, Dales S: **Biogenesis of vaccinia: interrelationship between post-translational cleavage, virus assembly, and maturation.** *Virology* 1982, **117**:341-356.
3. VanSlyke JK, Franke CA, Hruby DE: **Proteolytic maturation of vaccinia virus core proteins: identification of a conserved motif at the N termini of the 4b and 25K virion proteins.** *J Gen Virol* 1991, **72**:411-416.
4. VanSlyke JK, Whitehead SS, Wilson EM, Hruby DE: **The multistep proteolytic maturation pathway utilized by vaccinia virus P4a protein: a degenerate conserved cleavage motif within core proteins.** *Virology* 1991, **183**:467-478.
5. Byrd CM, Bolken TC, Hruby DE: **The vaccinia virus I7L gene product is the core protein proteinase.** *J Virol* 2002, **76**:8973-6.

6. Byrd CM, Bolken TC, Hruby DE: **Molecular dissection of the vaccinia virus I7L core protein proteinase.** *J Virol* 2003, **77**:11279-11283.
7. Ericsson M, Cudmore S, Shuman S, Condit RC, Griffiths G, Locker JK: **Characterization of ts16, a temperature-sensitive mutant of vaccinia virus.** *J Virol* 1995, **69**:7072-7086.
8. Kane EM, Shuman S: **Vaccinia virus morphogenesis is blocked by a temperature-sensitive mutation in the I7 gene that encodes a virion component.** *J Virol* 1993, **67**:2689-2698.
9. Ansarah-Sobrinho C, Moss B: **Role of the I7 protein in proteolytic processing of vaccinia virus membrane and core components.** *J Virol* 2004, **78**:6335-6343.
10. Byrd CM, Hruby DE: **A conditional-lethal vaccinia virus mutant demonstrates that the I7L gene product is required for virion morphogenesis.** *Virology* 2005, **2**:4.
11. Webster A, Russell S, Talbot P, Russell WC, Kemp GD: **Characterization of the adenovirus proteinase: substrate specificity.** *J Gen Virol* 1989, **70**:3225-3234.
12. Andres G, Alejo A, Simon-Mateo C, Salas ML: **African swine fever virus protease, a new viral member of the SUMO-1-specific protease family.** *J Biol Chem* 2001, **276**:780-787.
13. Li SJ, Hochstrasser M: **A new protease required for cell-cycle progression in yeast.** *Nature* 1999, **398**:246-51.
14. Byrd CM, Bolken TC, Mjalli AM, Arimilli MN, Andrews RC, Rothlein R, Andrea T, Rao M, Owens KL, Hruby DE: **New class of orthopoxvirus antiviral drugs that block viral maturation.** *J Virol* 2004, **78**:12147-12156.
15. Webster A, Hay RT, Kemp GD: **The adenovirus protease is activated by a virus-coded disulphide-linked peptide.** *Cell* 1993, **72**:97-104.
16. Webster A, Russell WC, Kemp GD: **Characterization of the adenovirus proteinase: development and use of a specific peptide assay.** *J Gen Virol* 1989, **70**:3215-3223.
17. Rubio D, Alejo A, Rodriguez I, Salas ML: **Polyprotein processing protease of African swine fever virus: purification and biochemical characterization.** *J Virol* 2003, **77**:4444-4448.
18. Raczynski P, Condit RC: **Specific inhibition of vaccinia virus growth by 2'-O-methyladenosine: isolation of a drug-resistant virus mutant.** *Virology* 1983, **128**:458-62.
19. Hruby DE, Guarino LA, Kates JR: **Vaccinia virus replication. I. Requirement for the host-cell nucleus.** *J Virol* 1979, **29**:705-15.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

